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Risk factors and molecular epidemiology of community-onset, multidrug resistance extended-spectrum β -lactamase-producing *Escherichia coli* infections

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Division of Infectious Diseases, Samsung Medical Center, Sungkyunkwan University School of Medicine, 81 Irwon-ro, Gangnamgu, Seoul 06351, Korea Tel: +82-2-3410-0324 Fax: +82-2-3410-0064 E-mail: collacin@hotmail.com **Background/Aims:** Although multidrug resistance (MDR) among extended-spectrum β -lactamase-producing *Escherichia coli* (ESBL-EC) poses significant therapeutic challenges, little is known regarding the risk factors and epidemiology of community-onset MDR-ESBL-EC infections. We performed this study to investigate risk factors and the molecular epidemiology of community-onset MDR-ESBL-EC infections.

Methods: We conducted a case-control-control study of community-onset infections. MDR-ESBL-EC was defined as ESBL-EC that demonstrated *in vitro* resistance to trimethoprim-sulfamethoxazole, fluoroquinolones (FQs), and gentamicin. Patients with MDR-ESBL-EC infections were designated as case patients. A control group I (CG I) patient was defined as a person whose clinical sample yielded ESBL-EC that did not meet the criteria for MDR. A control group II (CG II) patient was defined as a patient with a non-ESBL-EC infection.

Results: Of 108 patients with ESBL-EC infections, 30 cases (27.8%) were due to MDR-ESBL-EC. Compared with CG I, prior use of FQs (odds ratio [OR], 3.16; 95% confidence interval [CI], 1.11 to 8.98) and immunosuppressant use (OR, 10.47; 95% CI, 1.07 to 102.57) were significantly associated with MDR-ESBL-EC. Compared with CG II, prior use of FQs (OR, 15.53; 95% CI, 2.86 to 84.27) and healthcare-associated infection (OR, 5.98; 95% CI, 2.26 to 15.86) were significantly associated with MDR-ESBL-EC. CTX-M-15 was the most common in MDR-ESBL-EC infections (59.1% [13/22]), while CTX-M-14 was the most common in non-MDR-ESBL-EC infections (41.6% [32/77]). CTX-M-15 was significantly associated with MDR-ESBL-EC infections (59.1% vs. 32.5%, p = 0.028). Pulsed-field gel electrophoresis showed clonal diversity of MDR-ESBL-EC isolates.

Conclusions: The emergence of strains of MDR-ESBL-EC in the community poses an important new public health threat. More information on the emergence and transmission of these strains will be necessary in order to prevent their spread.

Keywords: Drug resistance, multiple; Extended-spectrum β -lactamase; Escherichia coli; Community-onset infection

INTRODUCTION

Extended-spectrum β-lactamase-producing Escherichia coli (ESBL-EC) have become increasingly prevalent nosocomial pathogens [1-3]. During recent years, community-onset infections due to ESBL-EC have emerged worldwide [4,5]. ESBL-EC are also frequently resistant to non- β -lactam antibiotics, such as fluoroquinolones (FQs), trimethoprim-sulfamethoxazole (TMP-SMX), and aminoglycosides [2,4]. FQs, cephalosporins, and TMP-SMX are recommended as empirical therapies for community-onset infection due to E. coli [6-8]. Most patients with community-onset infections caused by these organisms have urinary tract infections (UTIs) or intra-abdominal infections [5,9]. FQs and TMP-SMX may be regarded as the treatment of choice or as alternative antimicrobial therapy for UTIs or intra-abdominal infections due to ESBL-EC, if the bacteria are susceptible in vitro to these agents [4].

The clinical relevance of multidrug resistance (MDR) among ESBL-EC (i.e., those resistant to multiple other antibiotics or antibiotic classes in addition to the oxymino β -lactams) is of great concern because therapeutic options are severely limited [10]. As the frequency of infection caused by MDR-ESBL-EC is increasing, so the use of carbapenems is rising, which may contribute to the spread of carbapenem resistance. Moreover, isolation of carbapenem resistant ESBL-producing Enterobacteriaceae has been increasingly reported [11]. In addition, empirical antibiotic treatment of community-onset infections may be inappropriate if MDR-ESBL-EC are highly prevalent in the community.

Even though MDR-ESBL-EC poses significant therapeutic challenges, there is still little clinical information about community-onset MDR-ESBL-EC infections. Although several studies have reported on hospital-acquired MDR-ESBL-EC infections, data regarding community-onset MDR-ESBL-EC infections are limited [10,12,13].

This study was conducted to determine risk factors and the molecular epidemiology of MDR among ES-BL-EC causing community-onset infections. We wanted to identify risk factors associated with the acquisition of community-onset MDR-ESBL-EC infections. In addition, we wanted to identify risk factors associated with acquisition of a MDR-ESBL-EC among patients



with community-onset ESBL-EC infections. For this purpose, we employed the case-control-control study design, which enables more accurate identification of risk factors for multidrug-resistant pathogens than the standard case-control study design [14]. Using data collected in a 9-month period at two university hospitals, we conducted parallel analyses in which patients who presented with an infection caused by MDR-ESBL-EC were compared to patients who presented with an infection caused by non-MDR-ESBL-EC and patients with non-ESBL-EC infections.

METHODS

Study design and patients

We performed a post hoc analysis of individual patients included in a previous study [15]. In the current study, a case-control-control design was used to evaluate risk factors associated with MDR-ESBL-EC in community-onset infections. Study subjects were prospectively identified as previously described [15] through a computerized database maintained by the clinical microbiology laboratory. The study was conducted between September 2010 and May 2011 at Samsung Medical Center in Seoul (a 1,950-bed tertiary care university hospital) and at Samsung Changwon Hospital (a 700-bed community-based university-affiliated hospital in Changwon, Korea). Patients with community-onset infection due to E. coli were included in the study. The inclusion and exclusion criteria were the same as reported previously [15]. A case patient was defined as a person whose clinical sample yielded MDR-ESBL-EC. A control group I (CG I) patient was defined as a person whose clinical sample yielded ESBL-EC that did not meet the criteria for MDR. A control group II (CG II) patient was defined as a patient with a non-ESBL-EC infection. For each patient who presented with an infection caused by ES-BL-EC, one CG II was randomly chosen from a group of outpatients from whom non-ESBL-EC had been isolated from a clinical sample that had been sent to the same laboratory for culturing during the week following presentation of the case patient. The CG II patients were matched according to the culture specimen and acquisition unit. The patient data collected included age, sex, underlying disease, severity of underlying diseases as

classified by McCabe and Jackson criteria [16], presence of severe sepsis or septic shock at presentation, severity of illness at presentation according to the Pitt bacteremia score [2], and receipt of any antimicrobial therapy prior to onset of the infection. The presence of any of the following comorbid conditions was also documented: neutropenia, recent surgical procedure within the prior 3 months, corticosteroid use within the prior 1 month, immunosuppressive therapy within the prior 1 month, or the presence of an indwelling urinary catheter or percutaneous tube. Because the study was observational, the attending physician decided on indications for cultures, other tests, and treatments based on each patient's individual clinical situation. The study was approved by the Institutional Review Board of Samsung Medical Center, Seoul, Korea. Written informed consent was not required because of the observational nature of the study.

Definitions

The community-onset infections were further classified as healthcare-associated (HCA) or community-associated (CA). Episodes were considered to be HCA if any of the following criteria were present: a history of a 48 hours or longer hospital admission in the previous 90 days, hemodialysis, intravenous medication, home wound care in the previous 30 days, or residence in a nursing home or long-term care facility [9,17]. Community-onset infections that did not meet these criteria were considered to be CA. MDR-ESBL-EC was defined as an ESBL-EC that demonstrated in vitro resistance to all of the following three antibiotics or antibiotic classes: TMP-SMX, FQs (i.e., ciprofloxacin and levofloxacin), and gentamicin. Non-MDR-ESBL-EC was defined as ESBL-EC that did not meet the criteria for MDR. Community-onset infection was defined as an infection diagnosed within the first 48 hours of hospitalization. The sites of infection were determined by the physicians on the basis of clinical evaluation and the isolation of EC from the presumed portal of entry [9,18]. Primary bacteremia was defined according to Centers for Disease Control and Prevention definitions [18]. Patients with immunosuppression included those who were on immunosuppressive therapy (chemotherapeutic agents, immunosuppressive agents, or radiation therapy). Neutropenia was defined as an absolute neutrophil count < 500 neutrophils/mm³. Sepsis was defined as the presence of a systemic inflammatory response syndrome (any two of the following: tachypnea > 20 breaths per minute, white blood cell count < 4,000 or > 12,000 cells/ μ L, heart rate > 90 beats per minute, and fever > 38.0°C or hypothermia < 36.0°C) in addition to a documented or presumed infection [19]. Severe sepsis was defined as sepsis with one or more clinical signs of organ dysfunction and septic shock defined as sepsis with hypotension despite adequate fluid resuscitation along with the presence of perfusion abnormalities [19]. Prior antibiotic therapy was defined as the receipt of any systemic antibiotics for more than 48 hours in the preceding 30 days.

Microbiologic analysis

Identification of microorganisms, antimicrobial susceptibility testing, and ESBL confirmatory testing were performed as previously described [15]. Quality control was performed using the strains *E. coli* 25922 and Pseudomonas aeruginosa 27853.

Detection of ESBL genes and sequencing of the polymerase chain reaction product

For the characterization of ESBL types, polymerase chain reactions (PCRs) and sequencing of PCR products were performed using available stored isolates, as described previously [15]. ESBL-related genes, such as TEM, SHV, CTX-M, and OXA, were amplified by PCR from clinical isolates as described in previous studies [20,21]. The types of ESBL genes were identified by comparing the sequences to those in the database of G. Jacoby and K. Bush (http//lahey.org/Studies/).

Genetic typing by pulsed-field gel electrophoresis

Available MDR-ESBL-EC isolates were evaluated for genetic relatedness by pulsed-field gel electrophoresis (PFGE). For PFGE, agarose-embedded bacterial genomic DNA was digested with 20 U Xba l. The restriction fragments were separated by electrophoresis in 0.5 × Tris/borate/ethylenediaminetetraacetic acid (TBE) buffer. The subsequent PFGE analyses were performed on a CHEF-MAPPER XA apparatus (Bio-Rad Laboratories, Hercules, CA, USA) as described previously [22]. The isolates were considered genetically related if the Dice coefficient correlation was 80% or greater, which corresponds to the possibly related criterion of Tenover et al. [23].

Statistical analysis

The Student *t* test or the Mann-Whitney test was used to compare continuous variables, and the chi-square or Fisher exact test was used to compare categorical variables. We used a backward stepwise logistic regression analysis to control for the effects of confounding variables. All risk factors with a p < 0.1 at the bivariate level were included in the multivariate logistic model predicting MDR-ESBL-EC infection. All variables for which the *p* value was less than 0.05 in the multivariate analysis were retained in the final model. Interactions between variables were not introduced into the models. Odds ratios (ORs) and their 95% confidence intervals (CIs) were calculated. All *p* values were two-tailed, and *p* < 0.05 was considered statistically significant. IBM SPSS version 19 (IBM Co., Armonk, NY, USA) was used for these analyses.

RESULTS

Clinical characteristics and risk factors for community-onset infections caused by MDR-ESBL-EC

During the study period, a total of 108 patients with community-onset ESBL-EC infections were identified and all of them were included in this study. Of the 108 isolates, 30 (27.8%) were due to MDR-ESBL-EC. The mean age (± standard deviation) of the study population was 62.4 ± 16.0 years, and 70 (64.8%) were women. The most common underlying disease was diabetes mellitus (26.9% [28/108]) and the most common site of infection was the urinary tract (67.6% [73/108]). To analyze risk factors associated with community-onset MDR-ESBL-EC, data from the 30 patients with MDR-ESBL-EC were compared to those of the 78 patients with non-MDR-ESBL-EC (CG I) and 100 patients with non-ESBL-EC infections (CG II). The clinical characteristics of the case and control groups and risk factors for community-onset MDR-ESBL-EC infections are summarized in Table 1. Variables such as age, sex, underlying disease, and severity of illness at presentation were similar between the case and control groups. The source of isolates did not differ significantly between case and control groups. Compared with CG I, significant factors associated with community-onset MDR-ESBL-EC infections were immunosuppressive therapy and prior receipt of FQs (all p < 0.05). Compared with CG II, HCA infection, severity



of illness at presentation, immunosuppressive therapy, and prior receipt of antibiotics, especially FQs, were significantly associated with MDR-ESBL-EC infections.

To identify independent risk factors associated with community-onset MDR-ESBL-EC infections, a logistic regression analysis was performed including variables with an association of p < 0.1 at the univariate level. The independent risk factors associated with community-onset MDR-ESBL-EC infections are shown in Table 2. Immunosuppressive therapy (OR, 10.47; 95% CI, 1.07 to 12.05; p = 0.044 in model 1), prior receipt of FQs (OR, 3.16; 95% CI, 1.11 to 8.98; p = 0.31 in model 1; OR, 15.53; 95% CI, 2.86 to 84.27; p = 0.01 in model 2), and HCA infection (OR, 5.98; 95% CI, 2.26 to 15.86; p < 0.01 in model 2) were found to be independent factors associated with community-onset MDR-ESBL-EC infections.

Microbiological characteristics of MDR-ESBL-EC causing community-onset infections

The 108 community-onset ESBL-EC isolates demonstrated variable resistance to other antibiotics. The prevalence of resistance to piperacillin/tazobactam (36.7% vs. 29.5%, p = 0.472) and amoxicillin/clavulanic acid (73.3%) vs. 59.0%, p = 0.189) were similar for the MDR-ESBL-EC group and the non-MDR-ESBL-EC group. However, strains resistant to tobramycin were more frequent in the MDR-ESBL-EC group than the non-MDR-ESBL-EC group (93.3% vs. 44.9%, *p* < 0.001) (Table 3). None of the strains were resistant to imipenem or meropenem. Both MDR- and non-MDR-ESBL-EC isolates were highly susceptible to amikacin (100% vs. 98.7%, *p* = 1.000). Ninetynine isolates of ESBL-EC were available for further microbiological study. All of these isolates produced ESBLs from the CTX-M family. We compared MDR-ESBL-EC with non-MDR-ESBL-EC and the distribution of ESBL genes is summarized in Table 4. CTX-M-15 was the most common type of MDR-ESBL-EC (59.1% [13/22]), while CTX-M-14 was the most common non-MDR-ESBL-EC (41.6% [32/77]). CTX-M-15 was significantly associated with MDR-ESBL-EC (59.1% vs. 32.5%, *p* = 0.028).

PFGE of 22 MDR-ESBL-EC strains was performed to characterize the clonality of these isolates. Because two isolates (test number 1 and 19) among the 22 MDR-ES-BL-EC did not provide DNA bands by the PFGE method used in this study, we analyzed only 20 isolates. Applying a similarity index of 80% to the PFGE results, four



Table 1. Univariate analysis of risk factors for community-onset multidrug-resistant extended-spectrum β -lactamase-producing *Escherichia coli* infections

Risk factor	Case group (n = 30)	Control group I^a (n = 78)	p value	Control group II ^b (n = 100)	p value
Female sex	22 (73.3)	48 (61.5)	0.250	79 (79.0)	0.513
Older age (≥ 65 yr)	17 (56.7)	39 (50.0)	0.535	49 (49.0)	0.461
Healthcare-associated infection	20 (66.7)	36 (46.2)	0.056	20 (20.0)	< 0.001
Underlying disease					
Diabetes mellitus	8 (26.7)	20 (25.6)	0.913	24 (24.0)	0.766
Liver disease	4 (13.3)	9 (11.5)	0.752	10 (10.0)	0.737
Renal disease	6 (20.0)	8 (10.3)	0.206	15 (15.0)	0.573
Solid tumor	6 (20.0)	18 (23.1)	0.730	20 (20.0)	1.000
Neurologic disease	6 (20.0)	15 (19.2)	0.928	10 (10.0)	0.201
Severity of underlying illness by McCabe and Ja	ackson criteria	a			
Non-fatal	18 (60.0)	58 (74.4)	0.218	85 (85.0)	
Ultimately fatal	11 (36.7)	16 (20.5)		14 (14.0)	0.012
Rapidly fatal	1 (3.3)	4 (5.1)		1 (1.0)	
Comorbid condition					
Recent operation	1 (3.3)	5 (6.4)	1.000	3 (3.0)	1.000
Neutropenia	3 (10.0)	2 (2.6)	0.130	2 (2.0)	0.080
Immunosuppressive therapy	4 (13.3)	1 (1.3)	0.020	3 (3.0)	0.049
Central venous catheterization	2 (6.7)	4 (5.1)	0.669	3 (3.0)	0.326
Indwelling urinary catheter	1 (3.3)	7 (9.0)	0.439	5 (5.0)	1.000
Percutaneous tube	5 (16.7)	6 (7.7)	0.175	5 (5.0)	0.050
Prior receipt of antibiotics	13 (43.3)	26 (33.3)	0.333	8 (8.0)	< 0.001
Cephalosporin	4 (13.3)	14 (17.9)	0.564	4 (4.0)	0.082
β -Lactam or β -lactam/ β -lactamase inhibitor	1 (3.3)	7 (9.0)	0.439	1 (1.0)	0.410
Fluoroquinolone	10 (33.3)	10 (12.8)	0.014	2 (2.0)	< 0.001
Severity of illness					
No sepsis	20 (66.7)	51 (65.4)	0.611	66 (66.o)	0.671
Sepsis	5 (16.7)	18 (23.1)		22 (22.0)	
Severe sepsis	1 (3.3)	4 (5.1)		5 (5.0)	
Septic shock	4 (13.3)	5 (6.4)		7 (7.0)	
Site of infection					
Urinary tract	20 (66.7)	53 (67.9)	1.000	67 (67.0)	1.000
Pancreaticobiliary tract	2 (6.7)	10 (12.8)	0.504	11 (11.0)	0.731
Intra-abdomen	5 (16.7)	5 (6.4)	0.137	7 (7.0)	0.146
Unknown	2 (6.7)	3 (3.8)	0.616	3 (3.0)	0.326

Values are presented as number (%).

^aControl group I: those whose clinical sample yielded extended-spectrum β -lactamase-producing *Escherichia coli* (ESBL-EC) that did not meet the criteria for multidrug resistance.

^bControl group II: those with non-ESBL-EC infections.



Table 2. Independent risk factors for multidrug-resistance in community-onset infections caused by extended-spectrum
β-lactamase-producing Escherichia coli

Risk factor	Adjusted OR (95% CI)	p value				
Model 1 (with control group I)						
Immunosuppressive therapy	10.47 (1.07–102.57)	0.044				
Prior receipt of fluoroquinolone	3.16 (1.11–8.98)	0.031				
Model 2 (with control group II)						
Prior receipt of fluoroquinolone	15.53 (2.86–84.27)	0.001				
Healthcare-associated infection	5.98 (2.26–15.86)	< 0.001				

OR, odds ratio; CI, confidence interval.

Table 3. Comparison of rates of resistance to antimicrobials of Escherichia coli isolates causing community-onset infections^a

	% of patients (no. taking drug/total)					
Antibiotic	MDR-ESBL-EC group	Non-MDR- ESBL-EC group	p value	Non-ESBL-EC group	p value	
Amoxicillin-clavulanate	73.3 (22/30)	59.0 (46/78)	0.189	11.0 (11/100)	< 0.001	
Piperacillin-tazobactam	36.7 (11/30)	29.5 (23/78)	0.472	11.0 (11/100)	0.001	
Amikacin	0.0 (0/30)	1.3 (1/78)	1.000	0.0 (0/30)		
Gentamicin	100.0 (30/30)	38.5 (30/78)	< 0.001	20.0 (20/100)	< 0.001	
Tobramycin	93.3 (28/30)	44.9 (33/78)	< 0.001	16.0 (16/100)	< 0.001	
Trimethoprim- sulfamethoxazole	100.0 (30/30)	34.6 (27/78)	< 0.001	22.0 (22/100)	< 0.001	
Levofloxacin	100.0 (30/30)	71.8 (56/78)	< 0.001	26.0 (26/100)	< 0.001	

MDR, multidrug resistance; ESBL-EC, extended-spectrum β -lactamase-producing *Escherichia coli*. ^aAccording to automatic susceptibility testing.

Table 4. Comparison of ESBL types between MDR-ESBL-EC and non-MDR-ESBL-EC isolates

Variable	MDR-ESBL-EC (n = 22)	Non-MDR-ESBL-EC ($n = 77$)	p value
CTX-M-14	7 (31.8)	32 (41.6)	0.466
CTX-M-15	13 (59.1)	25 (32.5)	0.028
CTX-M-14 and CTX-M-15	2 (9.1)	1 (1.3)	0.123
Other CTX-M type ^a	0	19 (24.7)	0.006

Values are presented as number (%).

 $\label{eq:spectrum} \text{ESBL-EC}, extended-spectrum \ensuremath{\beta}\xspace - producing \ensuremath{\textit{Escherichia coli}}; \ensuremath{\textit{MDR}}\xspace, multidrug \ensuremath{\textit{resistance}}\xspace.$

^aOther CTX-M: CTX-M-24, 2 isolates; CTX-M-27, 10 isolates; CTX-M-3, 1 isolate; CTX-M-57, 6 isolates.

clonal clusters were identified, consisting of two or three isolates for each cluster. The 10 remaining isolates showed greater clonal diversity (Fig. 1). The microbiological characteristics of MDR-ESBL-EC that were available for further microbiological evaluation are shown in Table 5.

DISCUSSION

Our data showed that 27.8% of community-onset ES-BL-EC isolates were MDR and significant risk factors for MDR among ESBL-EC isolates causing community-onset infections were prior exposure to FQs, receipt of immunosuppressive therapy, and HCA infection. Previous

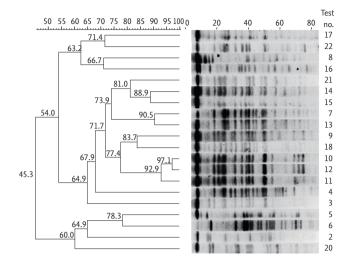


Figure 1. Dendrogram showing the relatedness of pulsed-field gel electrophoresis banding patterns for 20 multidrug resistance extended-spectrum β -lactamase-producing *Escherichia coli* isolates. Similarity among profiles was determined by cluster analysis in the dendrogram using a cutoff of at least 80%.

studies have suggested that the prevalence of community-onset ESBL-EC infections varies widely by geographic region, and most ESBL-producing isolates show co-resistance to multiple antibiotics [5,9,24,25]. However, data on risk factors for community-onset MDR-ESBL-EC infections are limited. Although two previous studies evaluated risk factors for infections by MDR-ESBL-EC, these studies included mainly nosocomial infections and the proportion of community-onset infections was less than 10% of the study population [10,13]. To the best of our knowledge, this is the first study to investigate risk factors and molecular epidemiology of MDR-ESBL-EC isolates among community-onset infections.

In the current study, we sought to identify risk factors for MDR-ESBL-EC among community-onset infections. The rate of MDR among ESBL-EC isolates was similar to rates found in previous studies [10,12]. We found that the receipt of immunosuppressive therapy, prior exposure to FQs, and HCA infection were associated with community-onset MDR-ESBL-EC infections. Patients with community-onset infection due to an MDR organism were more likely to be prescribed inactive empirical therapy. The MDR-ESBL-EC group received more inappropriate empirical therapy than the non-MDR-ESBL-EC group (90.0% vs. 66.7%, p = 0.015). Patients on immunosuppressive therapy, those with prior exposure to FQs, and those with HCA infections should be offered carbapenems as empirical therapy even for community-onset infections.

In our previous study, prior receipt of FQs was an independent risk factor for nosocomial MDR-ESBL-EC and Klebsiella pneumoniae infections [12]. In our previous studies, HCA infection and prior receipt of FQs were associated with community-onset ESBL-EC infections [15]. In other previous studies, FQ use has been associated with the acquisition of other MDR gram-negative infections, such as P. aeruginosa and Acinetobacter baumannii [26,27]. In addition, Villers et al. [27] showed that the institution of a policy restricting the use of intravenous FQs could decrease the rate of infection with A. baumannii. Interestingly, a recent study showed a correlation between FQ resistance and MDR in ESBL-EC infections [28]. Because this has been previously reported for MDR gram-negative infections [26,29,30], we can speculate that prior receipt of a FQ either results in selective pressure or facilitates the activation of intrinsic mechanisms that confer resistance to multiple antibiotic drug classes, such as the drug efflux mechanism.

In our study, immunosuppressive therapy was also associated with community-onset MDR-ESBL-EC infections. Immunosuppression presents an especially troublesome situation for critically ill patients as it has been identified as an independent risk factor for infectious morbidity and mortality [31]. Not surprisingly, a significantly greater proportion of immunosuppressed patients received antibiotic therapy during their treatment. Patients treated with immunosuppressive agents had more exposure to antibiotics than immunocompetent patients, which might be related to their acquiring MDR-ESBL-EC infections.

We observed CTX-M-15 was significantly associated with MDR-ESBL-EC (59.1% vs. 32.5%, *p* = 0.028). In recent studies, CTX-M-14 and CTX-M-15 were the predominant types of ESBLs, and CTX-M-15 was significantly associated with ST131 [32,33]. ST131 *E. coli* isolates had a trend toward greater nonsusceptibility to ciprofloxacin and cefepime [32,33]. Therefore, our data suggests that CTX-M-15 isolates show more MDR patterns than CTX-M-14.

We performed PFGE to examine the molecular epidemiology of community-onset MDR-ESBL-EC. Ten isolates among 20 MDR-ESBL-EC were not clonally



Test no.	Center	Department	Speci- men	Date of culture	Cefepime MIC	Piperacillin/ tazobactam MIC	Ciprofloxacin MIC	Ertapenem MIC	ESBL types
1 ^a	SMC	ED	Urine	2010-10-04	8	8/4	> 128	≤ 0.03	CTX-M-14
2	SMC	ED	Blood	2010-10-20	64	2/4	8	≤ 0.03	CTX-M-14
3	SMC	ED	Blood	2011-02-10	128	8/4	>128	0.06	CTX-M-15
4	SMC	ED	Blood	2011-02-15	>128	16/4	128	0.25	CTX-M-15
5	SMC	ED	Urine	2011-02-28	16	2/4	4	≤ 0.03	CTX-M-14
6	SMC	ED	Urine	2011-03-15	32	8/4	>128	0.06	CTX-M-15
7	SMC	ED	Blood	2011-04-16	4	4/4	32	≤ 0.03	CTX-M-14
8	SMC	OPD	Urine	2011-05-18	>128	8/4	64	0.12	CTX-M-15
9	SMC	OPD	Urine	2011-05-31	>128	4/4	64	0.12	CTX-M-15
10	SCH	ED	Blood	2010-10-09	>128	8/4	64	0.06	CTX-M-14, CTX-M-15
11	SCH	ED	Blood	2010-10-10	>128	8/4	> 128	≤ 0.03	CTX-M-15
12	SCH	OPD	Blood	2010-10-15	>128	16/4	64	≤ 0.03	CTX-M-14, CTX-M-15
13	SCH	OPD	Urine	2010-10-18	4	2/4	32	≤ 0.03	CTX-M-15
14	SCH	ED	Blood	2010-10-18	64	4/4	>128	≤ 0.03	CTX-M-15
15	SCH	ED	Blood	2010-12-11	128	2/42/4	64	0.06	CTX-M-15
16	SCH	OPD	Urine	2010-12-27	>128	32/4	>128	0.25	CTX-M-15
17	SCH	ED	Urine	2011-01-27	128	8/4	> 128	0.06	CTX-M-14
18	SCH	ED	Blood	2011-04-01	>128	64/4	> 128	0.06	CTX-M-15
19 ^a	SCH	ED	Urine	2011-04-07	64	8/4	> 128	≤ 0.03	CTX-M-14
20	SCH	ED	Urine	2011-04-20	>128	8/4	>128	≤ 0.03	CTX-M-15
21	SCH	ED	Ascites	2011-04-30	64	16/4	128	≤ 0.03	CTX-M-15
22	SCH	ED	Pus	2010-10-11	>128	128/4	> 128	0.5	CTX-M-15

Table 5. Microbiological characteristics of community-onset multidrug-resistant extended-spectrum β -lactamase-producing *Escherichia coli* available for further microbiological studies

 $MIC, minimum \ inhibitory \ concentration; \ ESBL, extended-spectrum \ \beta-lactamase; \ SMC, \ Samsung \ Medical \ Center; \ ED, \ Emergency \ Department; \ OPD, \ Out \ Patient \ Department; \ SCH, \ Samsung \ Changwon \ Hospital.$

^aThese isolates did not show any pulsed-field gel electrophoresis band.

related. The remaining 10 isolates were classified into four closely related groups. Taking the PFGE results into consideration, it is reasonable to assume that the MDR-ESBL-EC isolates identified in this study belong to diverse clones.

Numerous studies have demonstrated that most ES-BL-producing *Enterobacteriaceae* are resistant to multiple antibiotic classes. In addition to the agents included in our definition of MDR, we also noted a high prevalence of resistance to piperacillin/tazobactam and tobramycin among the MDR-ESBL-EC isolates (36.7% and 93.3%, respectively).

The types of infection due to MDR-ESBL-EC were generally those expected for *E. coli*, with UTIs predominating, followed by intra-abdominal or pancreaticobiliary tract infections. FQs, TMP-SMX, and aminoglycosides are commonly used in community-onset infections, and if the infection is not severe, can be additional treatment options for UTIs.

We found a high prevalence of community-onset MDR-ESBL-EC isolates that also demonstrated resistance to additional antibiotics. Of great concern is the

fact that one in four ESBL-EC isolates in this study was MDR. As the prevalence of MDR-ESBL-EC infections increases, inappropriate antibiotic treatment will also increase. A previous study demonstrated that increased mortality is not related to ESBL production but to the fact that the empirical therapies used were more frequently inappropriate among patients with ESBL-EC [5]. Reliance on carbapenems will continue to increase because other effective treatment options are increasingly limited. However, the increasing use of carbapenems has been paralleled by the rapid emergence of carbapenem resistance [11]. We suggest the empirical use of carbapenems only for patients with community-onset UTIs or intra-abdominal sepsis with any risk factor for MDR-ESBL-EC. However, other classes of antibiotics. especially amikacin and piperacillin/tazobactam, can be used for empirical therapy for patients with less severe infections caused by gram-negative bacilli, based on antimicrobial susceptibility results [34,35].

It has been shown previously that an ESBL-producing organism can be transferred to a hospital environment from the community [36]. Failure to consider the emergence of drug-resistant organisms in the community could undermine infection-control efforts in hospitals and render empirical antibiotic therapy inadequate. Interventions to limit the emergence of ESBL-producing isolates have traditionally focused on restricting certain antimicrobial agents, such as third-generation cephalosporins, associated with ESBL infections. Although such interventions are certainly an important component in efforts to control the emergence of ESBL-producing isolates, infection-control measures are also likely to be critical in interrupting the spread of such organisms.

There are several recognized limitations to our study. First, our study was observational, and thus, unknown risk factors might have been unequally distributed between the study groups. Harris et al. [37] suggested that using control patients infected with susceptible pathogens biases studies from the null hypothesis. Hence, we used a case-control-control study to assess the identifiable risk factors associated with community-onset MDR-ESBL-EC. Limitations related to the case-controlcontrol study design may also include the fact that the patients in CG II may not truly represent the population. We matched case and control patients on the basis of hospital and time period but intentionally did not

match for variables such as age, sex, or severity of underlying conditions, which might themselves have been risk factors [38]. Second, not all ESBL-EC isolates were available for further molecular analysis. Therefore, the ESBL characterization results may not be representative of all ESBL-EC isolates. However, there was no systematic process affecting which isolates were not available and more than 91% of isolates were available for molecular analysis. Third, plasmid analysis was not done. Thus, the possibility of dissemination of MDR-ESBL-EC isolates through the spread of such extrachromosomal genetic elements (even in the presence of unrelated PFGE patterns) cannot be excluded. Fourth, because previous exposure to FQs is likely to be associated with nosocomial acquisition, a well-established risk factor for resistance, our findings may have been affected by known or unknown confounders. Finally, our study was conducted in a large referral care medical center and a smaller community hospital. Thus, the results may not be generalizable to other types of institutions.

In conclusion, the emergence of MDR among community-onset ESBL-EC is particularly troublesome. We determined that 27.8% of ESBL-EC isolates, even in community-onset infections, were MDR and that the independent risk factors for infections caused by such isolates were prior exposure to FQs, receipt of immunosuppressive therapy, and HCA infection. Clinicians should be aware of the occurrence of community-onset MDR-ESBL-EC among predisposed patients and select initial appropriate empirical antimicrobial therapy to reduce morbidity and mortality in such patients. Given that a limited number of new antimicrobial agents are being developed to treat MDR-ESBL-EC, rigorous infection control measures are advocated to prevent the spread of this microorganism. Also, an assessment of the effectiveness of strategies implemented to minimize the exposure to FQs in patients at risk for developing community-onset ESBL-EC infections would be of interest.

KEY MESSAGE

 Among community-onset extended-spectrum β-lactamase-producing *Escherichia coli* (ES-BL-EC) infections, one in four ESBL-EC isolates was multidrug resistance (MDR).



2. The independent risk factors for communityonset MDR ESBL-EC infection were prior exposure to fluoroquinolones, receipt of immunosuppressive therapy, and healthcareassociated infection.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

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